



ORIGINALES

PEG-Interferon- α ribavirin-induced HCV viral clearance: a pharmacogenetic multicenter Spanish study

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Abstract

Objective: Dual PEGylated interferon- α (PEG-IFN) and ribavirin therapy has been the main hepatitis C virus (HCV) treatment of the last decade. Current direct-acting antiviral agents have improved the outcome of therapy but also have increased the cost and management complexity of treatment. The current study analyzes host genetics, viral and clinical predictors of sustained viral response (SVR) to dual PEG-IFN and ribavirin therapy in a representative Spanish population.

Methods: Observational prospective multicentre pharmacogenetic cohort study conducted in 12 different hospitals of 12 different Spanish regions. A total of 98 patients with SVR and 106 with non-SVR in response to PEG-IFN and ribavirin therapy were included. 33 single nucleotide polymorphisms located in 24 different genes related with inflammatory, immune and virus response were selected. Clinical and viral data were also analyzed as candidate of SVR predictors.

Results: IL-28B (rs12979860, rs7248668, rs8105790, rs8099917) and TNFRSF1B (rs1061622) genotypes, as well as TNFRSF1B/IL-10/TNF α (-308) non-TTG and TNFRSF1B/IL-10/IL-4 non-TTC haplotypes together with lower age, lower basal HCV RNA load, higher basal serum LDL cholesterol va-

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Resumen

Objetivo: El interferón- α pegilado (IFN-PEG) junto a ribavirina ha sido el principal tratamiento de la infección por el virus de la hepatitis C (VHC) de la última década. Los agentes antivirales de acción directa actuales han mejorado los resultados de la terapia, pero también han aumentado el costo y la gestión de la complejidad del tratamiento. El presente estudio analiza factores genéticos de los pacientes, así como predictores virales y clínicos de respuesta sostenida viral (RSV) al tratamiento con IFN-PEG y ribavirina en población Española.

Métodos: Estudio farmacogenético, multicéntrico, prospectivo, observacional de cohortes realizado en 12 hospitales diferentes de 12 comunidades autónomas diferentes. Se incluyeron un total de 98 pacientes con RVS y 106 sin SVR al tratamiento con IFN-PEG y ribavirina. Se seleccionaron 33 polimorfismos de nucleótido único ubicados en 24 genes diferentes relacionados con la respuesta inflamatoria, inmunológica y viral. Los datos clínicos y virales también se analizaron como candidatos predictores de RVS.

Resultados: Los genotipos IL-28B (rs12979860, rs7248668, rs8105790, rs8099917) y TNFRSF1B (rs1061622), así como los haplotipos TNFRSF1B / IL-10 / TNF α (-308) no-TTG y TNFRSF1B

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lues, VHC genotypes 2 and 3 and basal low grade fibrosis 0-2 were associated with a SVR in the univariate analysis. Independent predictors of SVR in the multivariate analysis were IL-28B rs12979860 CC, TNFRSF1B/IL-10/IL-4 non-TTC along with low baseline HCV RNA load and HCV genotypes 2 and 3.

Conclusions: IL-28B rs12979860 CC, TNFRSF1B/ IL-10/ IL-4 non-TTC haplotype, low baseline HCV RNA load and HCV genotypes 2 and 3 may help to predict successful outcome to PEG-IFN/ribavirin therapy in Spanish population.

KEYWORDS

Hepatitis C; Pharmacogenetics; Sustained viral response; TNFRSF1B

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Introduction

Hepatitis C virus (HCV) infection is a major global health problem affecting approximately 170 million people worldwide¹. The prevalence varies notably from nearly 0.1%-1% in Europe until the 5% in Asia and Africa^{2,3}. More than 20 % of these patients will progress to cirrhosis, hepatocellular carcinoma, liver transplantation or death⁴. Therefore, all patients are candidates for antiviral therapy. Dual PEGylated interferon α (PEG-IFN) ribavirin therapy has been the main HCV treatment of the last decade. However, the percentage of patients achieving a sustained virologic response (SVR), defined as undetectable HCV RNA 24 weeks after completion of treatment, is below target especially for the difficult to treat HCV genotypes 1 and 4 with dual therapy. Recently, a number of direct-acting antiviral agents (DAAs) have been developed for use with PEG-IFN/ribavirin as triple or IFN-free therapies. The first DAAs generation boceprevir and telaprevir in combination with PEG-IFN/ribavirin increased the SVR rates to 66%-75% in previously untreated HCV1 adults vs. 38% in the PEG-IFN/ribavirin group^{5,6}. Although triple therapy has improved SVR rates, these combinations have some drawbacks, including drug-drug interactions, a low efficacy in patients with HCV 2, 3 and 4 genotypes and in those who have not responded to previous dual PEG-IFN/ribavirin therapy. In this respect, second-generation protease inhibitors, such as simeprevir, asunaprevir, and danoprevir, are currently being evaluated or used in combination with PEG-IFN/ribavirin in an effort to overcome the limited efficacy of the first-generation protease inhibitors^{7,9}. In addition, new inhibitors of the NS5A/NS5B HCV polymerase ledipasvir and sofosbuvir are currently available as part of PEG-IFN/ribavirin triple therapy as well as in IFN-free dual therapy¹⁰.

Understanding the host and viral factors associated with viral clearance is necessary for individualizing therapy to maximize SVR rates, prevent progression to liver disease, and increase the overall benefits of therapy with respect to its prohibitive costs.

/ IL-10 / IL-4 no-TTC junto con la menor edad, menor carga de ARN-VHC basal, valores elevados de colesterol LDL en suero basal, genotipos VHC2 y 3 y bajo grado de fibrosis basal (0-2) se asociaron con una RVS en el análisis univariante. Los predictores independientes de RVS en el análisis multivariante fueron el genotipo IL-28B rs12979860 CC, el haplotipo TNFRSF1B / IL-10 / IL-4 no-TTC junto con los bajos niveles basales de VHC-ARN y los genotipos virales VHC2 y 3.

Conclusiones: El genotipo IL-28B rs12979860 CC, el haplotipo TNFRSF1B / IL-10 / IL-4 haplotipos no-TTC, la carga viral basal baja y los genotipos del VHC2 y 3 pueden ayudar a predecir una buena respuesta a la terapia con IFN-PEG y ribavirina en población española.

PALABRAS CLAVE

Hepatitis C; Farmacogenética; Respuesta viral sostenida; TNFRSF1B

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The SVR rate varies drastically among different races and ethnicities, with patients of African ancestry having much lower response rates as compared with Caucasians and Asians, supporting genetic predisposition to response¹¹. Both host and viral factors have been previously reported to be associated with SVR. Predictors of SVR include HCV genotypes 2 and 3, lower baseline serum HCV RNA level, younger age, female sex, lower hepatic fibrosis stage, lack of insulin resistance, and lower body mass index¹². Host genetics is increasingly recognized as important modulator of the therapy success. Genome-wide association studies have shown that single nucleotide polymorphisms (SNPs) in or near the interleukin-28B (IL-28B) gene are significantly associated with the treatment outcome for HCV-1¹³. Therefore, the assessment of the host IL28B genotype is increasingly used among HCV genotype-1 patients to inform clinical prediction of treatment outcome¹⁴. More specifically, the rs12979860 and rs8099917 SNPs were the two most examined: they were found in linkage disequilibrium each other, and to be similarly informative as host predictors of sustained viral clearance. In a recent meta-analysis of results from nine studies regarding the correlation of IL28B genotype with PEG-IFN/ribavirin SVR in 3110 Caucasian HCV-1 patients, the favourable IL28B genotype CC was present in 41% of individuals, and 67% of them achieved SVR compared to 37% of those with the unfavourable genotypes¹⁵. Other pharmacogenetic studies have been performed in different populations, showing a broad number of SNPs located in different genes of the host immune response, inflammatory, fibrosis and virus response pathways to be associated with the SVR to PEG-IFN/ribavirin dual therapy¹⁶. Although genetic markers are used as predictors of PEG-IFN/ribavirin in clinics, these only explain the 60-70% of SVR in Caucasian population. Given the recent introduction of multiple new DAAs, HCV treatment options have increased in parallel to its elevated costs, making difficult the optimal treatment decision. Pharmacogenetic studies of specific

populations together with the analysis of different clinical variables may be of potential value to appropriately select candidates for dual PEG-IFN/ribavirin therapy in the era of new DAAs.

The present study was designed to analyze the predictive value of different SNPs located at different host immune, inflammatory and liver fibrotic genes in a representative multicenter Spanish cohort of responders and non-responder HCV patients treated with PEG-IFN/ribavirin therapy.

Methods

Study Design and Population

This is an observational prospective multicentre pharmacogenetic cohort study conducted in 12 different hospitals of 12 different Spanish regions (Valencia, Alicante, Cartagena, A Coruña, Córdoba, Navarra, Salamanca, Lleida, Barcelona, Madrid, Tenerife, Las palmas de Gran Canaria) considered as a representative national population. Each hospital recruited 10 VHC patients who reached SVR and 10 VHC patients who showed no response to double PEG-IFN/ribavirin therapy reaching a total of 240 patients. The follow up study period was between February 2010 and June 2013, and included 24 weeks of follow up after the end of PEG-IFN/ribavirin therapy. Inclusion criteria were: 1) older than 18 years, 2) HCV 1-4 positive diagnosis, 3) PEG-IFN/ribavirin dual therapy, 3) clinical data available for liver enzymes, lipid profile, drug adherence, and VHC viral load at baseline, and every 12 weeks until the end of follow up, 5) stored whole blood samples for genetic determinations. Exclusion criteria were: 1) Patients who interrupted treatment because of adverse events or who voluntarily dropped out, 2) patients who infected with hepatitis B virus or human immunodeficiency virus were excluded, 3) loss of more than 20% of clinical data referred in inclusion criteria, 4) loss of more than 20% of genotyping data.

All HCV infected patients enrolled in this investigation agreed to donate a blood sample for genotyping, after having signed an informed consent. This study was approved by the Ethical Committee of the General University Hospital (Valencia), Complejo Hospitalario Universitario A Coruña (Galicia), Reina Sofía Hospital (Córdoba), General Hospital (Alicante), University Clinical Hospital (Salamanca), University Clinic (Navarra), General University Hospital of Santa Lucía (Cartagena), Gregorio Marañón Hospital (Madrid), University Hospital of Bellvitge (Catalunya), University Hospital Arnau de Vilanova (Lleida), University Hospital Complejo INSULAR Materno-Infantil (Las Palmas de Gran Canaria) and University Hospital Nuestra Señora de Candelaria (Tenerife), and conducted according to provisions of the Declaration of Helsinki and Good Clinical Practice Guidelines.

Treatment regimens and definition of response

PEG-IFN alfa-2a or alfa-2b was administered at doses of 180 μ g or 1.5 μ g/kg once per week, respectively, in combination with weight-adjusted ribavirin (1000 mg/day for <75 kg and 1200 mg/day for \geq 75 kg). The scheduled treatment duration was 24 weeks for HCV2 and HCV3 genotype patients and 48 weeks for HCV1 genotype patients. SVR was defined as undetectable plasma HCV RNA 24 weeks after the completion of therapy. A decline

of plasma HCV RNA of less than 2 log₁₀ at week 12 or lack of undetectable HCV RNA at treatment week 24 after having presented an at least 2 log₁₀ decline but without reaching undetectability was considered as non-response.

Selection and genotyping of single-nucleotide polymorphisms and HCV RNA quantification

Genomic DNA was isolated from whole blood samples using the QIAamp DNA Blood Maxi Kit (Qiagen GmbH, Hilden, Germany). Selected SNPs included SNPs belonging to genes involved in the immune host response, inflammatory pathways, liver function and fibrosis and inflammatory cell signaling previously associated with SVR or with liver inflammation/ fibrosis in HCV-infected patients (see supplementary table 1)^{16,17}.

All SNPs were genotyped using allelic discrimination technique by means of real time PCR analysis in a 7900HT Fast Real-Time PCR System (Applied Biosystem, Foster City, CA, USA) using TaqMan® GTXpress™ Master Mix (Applied Biosystems) and 5'exonuclease TaqMan genotyping predesigned assays according to manufacturer's instructions (Applied Biosystems). HCV genotyping was performed using the Inno-LiPA assay (Innogenetics, Zwijndrecht, Belgium). Serum HCV RNA was quantified by reverse transcription-PCR, using the Cobas Amplicor HCV Monitor Test, v 2.0 (Roche, Basel, Switzerland).

Data analysis

A descriptive analysis of the baseline variables was conducted. Before statistical analysis, normality distribution and homogeneity of the variables were tested by the Kolmogorov-Smirnov test. Continuous variables were expressed as mean \pm SD or median (interquartile range), depending on its distribution, and discrete variables were expressed as percentage. Haploview V4.1 Software (<https://www.broad.harvard.edu/haploview/haploview>) was used to calculate the Hardy-Weinberg equilibrium for all SNPs using the χ^2 goodness-of-fit test. Those gene SNPs with a minor frequency allele less than 5% or a *P*-value of the Hardy-Weinberg equilibrium below 0.001 were excluded from the analysis.

Those SNPs that were associated with SVR showing a *P*-value less than 0.05 were selected for further analysis. The linkage disequilibrium values between the genetic

Supplementary Table 1: *Single nucleotide polymorphisms (SNPs) selected for genotyping*

Name/ Gene	Function	SNPs (rs)	ObsHET	PredHET	HWpval	MAF	Alleles	Excluded genotyping
CYP3A5*3	Liver inflammation	rs776746	0.113	0.116	1.0	0.062	C:T	
MDR1	Liver inflammation	rs1045642	0.483	0.482	1.0	0.404	G:A	
UGT1A9:-2152	Liver inflammation	rs17868320	0.177	0.162	0.3643	0.089	C:T	
UGT1A9:-275	Liver inflammation	rs6714486	0.177	0.272	1.61E-05	0.163	T:A	*HW P-value < 0.001
UGT1A9*3	Liver inflammation	rs72551330	0.034	0.034	1.0	0.017	T:C	#MAF <0.05
UGT1A8*3	Liver inflammation	rs17863762	0.054	0.053	1.0	0.027	G:A	#MAF <0.05
UGT2B7*2	Liver inflammation	rs7439366	0.512	0.494	0.7165	0.443	C:T	
GNB3	Liver inflammation	rs5443	0.517	0.47	0.2067	0.377	C:T	
PXR	Liver inflammation	rs3814055	0.438	0.462	0.5405	0.362	C:T	
IL-28B	Response/ Liver inflammation	rs12979860	0.478	0.467	0.8919	0.372	C:T	
IL-28B	Response/ Liver inflammation	rs7248668	0.305	0.395	0.0025	0.271	C:T	
IL-28B	Response/ Liver inflammation	rs8105790	0.305	0.376	0.0133	0.251	T:C	
IL-28B	Response/ Liver inflammation	rs8099917	0.31	0.374	0.0262	0.249	T:G	
ITPA	Ribavirin toxicity	rs1127354	0.089	0.103	0.2078	0.054	C:A	
HIF-1	Liver fibrosis	rs11549465	0.187	0.193	0.8438	0.108	C:T	
VEGR-2	Liver fibrosis	rs1870377	0.365	0.39	0.4183	0.266	T:A	
FGF2	Liver fibrosis	rs2922979	0.438	0.437	1.0	0.323	C:G	
IL-6R	Response/ Liver fibrosis/ inflammation	rs2228145	0.507	0.486	0.6545	0.416	A:C	
IL-6	Response/ Liver fibrosis/ inflammation	rs1800796	0.148	0.145	1.0	0.079	G:C	
MTHFR	Liver inflammation	rs1801133	0.443	0.483	0.2839	0.409	C:T	
MTHFR	Liver inflammation	rs1801131	0.365	0.408	0.1671	0.286	A:C	
SLCO1B1	Liver inflammation	rs11045879	0.261	0.262	1.0	0.155	T:C	
SLCO1B1	Liver inflammation	rs4149081	0.261	0.369	1.0E-4	0.244	G:A	*HW P-value < 0.001
TNF α	Response/ Liver inflammation	rs1799724	0.143	0.141	1.0	0.076	C:T	
TNF α	Response/ Liver inflammation	rs1800629	0.261	0.234	0.1721	0.135	G:A	
TNF α	Response/ Liver inflammation	rs361525	0.192	0.182	0.7424	0.101	G:A	

Supplementary Table 1 (cont.). Single nucleotide polymorphisms (SNPs) selected for genotyping

Name/ Gene	Function	SNPs (rs)	ObsHET	PredHET	HWpval	MAF	Alleles	Excluded genotyping
IL-10	Response/ Liver inflammation	rs1800896	0.626	0.5	0.057	0.495	C:T	
IL-4	Response/ Liver inflammation	rs2070874	0.291	0.307	0.5459	0.19	C:T	
TNFRSF1B	Liver inflammation/ fibrosis	rs1061622	0.399	0.358	0.1597	0.234	T:G	
FCGR3A	Immunity	rs396991	0.453	0.449	1.0	0.34	A:C	
FCGR2A	Immunity	rs1801274	0.517	0.5	0.7423	0.485	A:G	
STAT4	Inflammatory signalling	rs7574865	0.365	0.4	0.2678	0.276	G:T	
PTPN22	Inflammatory signalling	rs2476601	0.084	0.08	1.0	0.052	G:A	

*SNPs with Hardy-Weinberg-Equilibrium (HW) P-value <0.001; **SNPs with minor allele frequency (MAF) <0.05; ObsHET: Observed heterozygosity; PredHET: Predicted heterozygosity.

markers studied (measured in Lewontin's standardized disequilibrium coefficient D'), haplotype frequencies and haplotype-based association analyses were calculated using Haploview V4.1. Student's T test or Mann-Whitney U test was used to compare normally distributed continuous variables with every type of virological response (SVR and non-SVR). Comparisons of qualitative variables, including genotype, allele frequencies, clinical, analytical and therapy variables, with the different types of virological response were analyzed by the χ^2 test. Odds ratios (OR) and confidence intervals (CI 95%) were calculated using Woolf approximation. A multivariate logistic regression analysis, including as covariables age, HCV genotype as well as those factors that were associated with SVR with a P -value less than 0.05 in the univariate analysis, was conducted. Descriptive analyses were carried out using the SPSS statistical software package release 20.0 (IBM Corporation, Somers, New York, USA).

Results

Patients

A total of 240 patients fulfilled the inclusion criteria for this study. Of these, 36 (15%) individuals were excluded because the loss of more than 20% of clinical data (18 patients), loss of more than 20% of genotyping data (7 patients) or due to the blood coagulation of sample (11 patients). Finally, 204 patients were included in the final analysis. After the end of follow-up, 98 (48 %) showed a SVR and 106 (52 %) showed a non-SVR to dual PEG-IFN/ribavirin therapy. Clinical characteristic of patients with SVR and non-SVR are showed in table 1. The cohort of patients with SVR showed significant lower age (46.59 (44.7-48.4) vs. 52.08 (50.2-53.8) years old; $P < 0.05$),

lower basal HCV RNA load (5.76 ± 0.09 vs. 6.14 ± 0.08 log IU/mL; $P < 0.05$) and higher basal serum LDL cholesterol values (114.1 ± 38.7 vs. 95.3 ± 34.2 mg/dL; $P < 0.05$) compared with non-SVR patients. Furthermore the VHC genotypes 2 and 3 (OR (95% CI): 4.86 (2.14-10.24) $P = 3.25e-06$) and basal low grade fibrosis 0-2 (OR (95% CI): 3.59 (0.82-6.74) $P = 0.0051$) were associated with a SVR (table 1). No other differences were found between clinical characteristics of patients with SVR and non-SVR.

Single-nucleotide polymorphisms selection and association with SVR to PEG-IFN/ribavirin treatment

A total of 33 SNPs located in 24 different genes related with inflammatory response, immune response, response to virus and liver fibrosis were selected for the primary analysis (supplementary table 1). Of these, 4 SNPs were excluded as a result of either a minor allele frequency below 0.05 (2 SNPs), or a P -value of the Hardy-Weinberg equilibrium below 0.001 (2 SNPs). Other 29 SNPs were in Hardy-Weinberg equilibrium indicating that no population stratification bias or genotyping error existed.

After distribution analysis of different SNPs between patients with SVR and non-SVR we only observed a significant association with the SVR in the IL-28B gene (rs12979860, rs7248668, rs8105790, rs8099917), TNFRSF1B (rs1061622) and UGT1A9*3 (rs72551330) as represented in supplementary table 2 and table 2. In the last case, the UGT1A9*3 rs72551330 was discarded because of the low prevalence of its favorable genotype TC (3.4%).

The prediction power for SVR differed among the five SNPs: the CC homozygous genotype of IL28B rs12979860 SNP was more strongly associated (OR

Table 1. Main basal demographic and clinical characteristics of the groups studied

Basal clinical data	SVR (n=98)	Non-SVR (n=106)	χ^2	OR	OR (95%CI)	P value
Age (years)	46.59 (44.7-48.4)	52.08 (50.2-53.8)				0.001
Sex (male vs. female)	67/35	68/40	0.309	1.083	0.82-1.43	0.578
Weight (kg)	75.71 ± 18.5	74.5 ± 13.6				0.736
Genotype 1a/ 1b and 4 vs. 2 and 3. N/N	70/24	99/7	23.04	4.86	2.14-10.24	3.25e-06
Plasma HCV-RNA (log UI/mL)	5.76 ± 0.09	6.14 ± 0.08				0.0003
AST (IU/L)	69.11 ± 53.7	91.4 ± 99.7				0.0662
ALT (IU/L)	107.5 ± 86.7	107.4 ± 89.4				0.8740
GGT (IU/L)	123.2 ± 262.9	136.4 ± 155.5				0.3121
Cholesterol (mg/dl)	175 ± 39.3	169.4 ± 32.3				0.2635
LDL cholesterol (mg/dl)	114.1 ± 38.7	95.3 ± 34.2				0.0031
triglycerides (mg/dl)	114.2 ± 68.01	119.2 ± 69.2				0.6061
Liver fibrosis. F0-F2 vs. F3-F4. N/N	46/16	36/45	14.72	3.59	0.82-6.74	0.0051

Abbreviations: SNP, single nucleotide polymorphism; SVR, sustained virological response; OR, odds ratio; CI, confidence interval. Comparisons of qualitative variables with the different types of virological response were analyzed by the χ^2 test. Mann-Whitney U test was used to compare normally distributed continuous variables with every type of virological response. Odds ratios and 95% CI per genotype were estimated by applying unconditional logistic regression. The P values less than 0.05 were considered significant. Values are expressed as mean ± SD or mean (interquartile range).

(95% CI): 2.27 (1.57-3.27) $P=8.75e-08$ for CC vs. CT/TT) than the CC genotype of rs7248668 (OR (95% CI): 1.66 (1.26-2.18) $P=6.00e-04$ for CC vs. CT/TT), the TT genotype of rs8105790 (OR (95% CI): 1.69 (1.27-2.17) $P=7.00e-04$ for TT vs. TC/CC), the TT genotype of rs8099917 (OR (95% CI): 1.61 (1.23-2.09) $P=0.0016$ for TT vs. TG/GG) or the TNFRSF1B GG genotype of rs1061622 (OR (95% CI): 1.78 (1.27-2.51) $P=7.00e-04$ for GG vs. TG/TT) as show table 2.

HCV active infection is accompanied by chronic liver inflammation which may be monitored by simple hepatic enzyme serum analysis. In this work, both cohorts of responders and non-responder patients showed similar elevated aspartate transaminase (AST) and alanine aminotransferase (ALT) hepatic enzymes (table 1). Host genetic SNPs located in IL28B (rs12979860) and in GNB3 (rs5443) were associated with AST and ALT normalization on week 12 of treatment independently of the final response to treatment (table 3) suggesting a positive influence on the hepatic anti-inflammatory role of the PEG-IFN/ribavirin treatment.

Haplotype analysis among the responders and non-responders HCV patients

After analysing the linkage disequilibrium among the polymorphisms studied, only a moderate association of

IL28B rs12979860 with an IL28B rs7248668, rs8105790 and IL28B rs8099917 ($D' < 0.8$) and strong association of IL28B rs7248668, rs8105790 and IL28B rs8099917 ($D' < 0.8$; Figure 1) were found. The reconstructed haplotypes of the IL-28B polymorphisms did not improved the predictive value of rs12979860 (table 4) suggesting an independent predictive value for each IL-28B SNP. Next analysis was designed to detect potential gene-gene interactions between combinations of 2 or 3 different SNPs. Table 5 shows the association of different gene-gene interactions with the SVR to dual PEG-IFN/ribavirin therapy. Different inflammatory and immune response pathways such as the IL-10/IL-4 and IL-10/IL-6R haplotypes as well as interactions with the TNFRSF1B gene with IL-10, IL-4 and TNF α genes were significantly associated with SVR (table 5).

Multivariate analysis

According to the univariate analysis, SNPs in IL-28B (rs12979860, rs7248668, rs8105790, rs8099917) and TNFRSF1B (rs1061622) genes, as well as the haplotypes TNFRSF1B/IL-10/TNF α (-308) TTG and TNFRSF1B/IL-10/IL-4 TTC, were entered into a multivariate logistic regression model adjusted for age, HCV genotype, presence of advanced fibrosis, HCV baseline viral load and basal LDL cholesterol values. In the multivariate analysis TNFR-

Supplementary table 2. Genotype frequencies of single nucleotide polymorphisms in HCV infected PEG-IFN α -ribavirin therapy patients with and without SVR

Gene/ SNPs	Study cohort n=204 N (%)	SVR n= 98 (48 %) N (%)	Non-SVR n= 106 (52 %) N (%)	χ^2	OR	OR (95%CI)	P
IL-28B							
rs12979860							
CC	78 (38.2)	56 (71.8)	22 (28.2)	26.47	2.27	1.57-3.27	8.75e-08
CT	97 (47.5)	38 (39.2)	59 (60.8)				
TT	29(13.7)	4 (14.3)	25 (85.7)				
rs7248668							
CC	117 (52.0)	69 (59)	48 (41)	13.5	1.66	1.26-2.18	6.00e-04
CT	66 (30.9)	23 (34.8)	43 (65.2)				
TT	21 (9.8)	6 (28.6)	15 (71.4)				
rs8105790							
TT	118 (52.2)	70 (59.3)	48 (40.7)	13.02	1.69	1.27-2.17	7.00e-04
TC	65 (30.7)	22 (33.8)	43 (66.2)				
CC	21 (9.8)	6 (28.5)	15 (71.4)				
rs8099917							
TT	119 (58.3)	70 (58.8)	49 (41.2)	9.92	1.61	1.23-2.09	0.0016
TG	65 (31.4)	21 (32.3)	44 (67.7)				
GG	20 (9.3)	7 (35)	13(65)				
CYP3A5*3							
rs776746 (T6986C)							
CC	178 (87.3)	85 (47.8)	93 (52.2)	0.043	1.094	0.46-2.56	0.83
CT	23 (11.3)	11 (47.8)	12 (52.2)				
TT	1 (0.5)	1 (100)	0				
MDR1							
rs1045642 (C3435T)							
TT	33 (16.2)	13 (39.4)	20 (60.6)	1.34	1.024	0.56-2.14	0.41
CT	98 (48.0)	50 (51)	48 (49)				
CC	73 (35.3)	36 (48.6)	37 (51.4)				
UGT1A9							
rs17868320 (-2152)							
CC	165 (80.4)	78 (47.3)	87 (52.7)	0.014	1.045	0.51-2.13	0.9
CT	39 (18.1)	19 (48.7)	20 (51.3)				
rs6714486 (-275)							
AA	16 (6.4)	7 (43.7)	9 (56.2)	0.273	1.18	0.62-2.26	0.61
TA	40 (18.1)	17 (42.5)	23 (57.5)				
TT	148 (71.1)	71 (47.9)	77 (52.1)				
UGT1A9*3							
rs72551330							
TC	7 (3.4)	6 (85.7)	1 (14.3)	4.017	1.817	1.29-2.54	0.036
TT	196 (96.1)	92 (46.9)	104 (53.1)				

Supplementary table 2 (cont.). Genotype frequencies of single nucleotide polymorphisms in HCV infected PEG-IFN α -ribavirin therapy patients with and without SVR

	Study cohort n=204	SVR n= 98 (48 %)	Non-SVR n= 106 (52 %)				
UGT1A8*3							
rs17863762							
GA	15 (5.9)	9 (60)	6 (40)	0.695	0.607	0.18-1.98	0.405
GG	188 (90.7)	86 (45.8)	102 (54.2)				
UGT2B7*2							
rs7439366							
CC	60 (29.4)	26 (43.3)	34 (56.7)	1.334	0.981	0.671-1.86	0.513
CT	105 (51.5)	51 (48.6)	54 (51.4)				
TT	38 (18.6)	21 (55.3)	17 (44.7)				
GNB3							
rs5443 (C825T)							
CC	75 (36.8)	37 (49.3)	38 (50.7)	1.273	1.01	0.751-1.24	0.529
CT	104 (51.0)	52 (50)	52 (50)				
TT	24 (11.8)	9 (37.5)	15 (62.5)				
PXR							
rs3814055 (C-25385T)							
CC	85 (41.7)	46 (54.1)	39 (45.9)	2.271	0.95	0.784-1.28	0.321
CT	89 (43.6)	38 (42.7)	51 (57.3)				
TT	29 (14.2)	14 (48.3)	15 (51.7)				
ITPA							
rs1127354							
AA	3 (1)	1 (33.3)	2 (66.6)	0.588	1.47	0.546-3.96	0.443
CA	18 (8.3)	6 (33.3)	12 (66.6)				
CC	182 (88.7)	88 (48.4)	94 (51.6)				
HIF-1							
rs11549465 (C1772T)							
CC	161 (78.9)	79 (49.1)	82 (50.9)	2.844	1.04	0.742-1.981	0.241
CT	39 (19.1)	19 (48.7)	20 (51.3)				
TT	3 (1.5)	0 (0)	3 (100)				
VEGR-2							
rs1870377 (A-1719T)							
AA	17 (7.8)	4 (23.5)	13 (76.5)	3.764	1.14	0.57-2.14	0.152
TA	74 (35.8)	35 (47.3)	39 (52.7)				
TT	113 (54.9)	58 (51.3)	55 (48.7)				
FGF2							
rs2922979 (C754G)							
CC	92 (45.1)	43 (46.7)	49 (53.3)	0.942	1.13	0.77-2.41	0.418
CG	90 (44.1)	42 (46.7)	48 (53.3)				
GG	21 (10.3)	13 (61.9)	8 (38.1)				

Supplementary table 2 (cont.). Genotype frequencies of single nucleotide polymorphisms in HCV infected PEG-IFN α -ribavirin therapy patients with and without SVR

	Study cohort n=204	SVR n= 98 (48 %)	Non-SVR n= 106 (52 %)				
IL-6R							
rs2228145 (A1510C)							
AA	67 (32.8)	30 (44.8)	37 (55.2)	0.851	1.09	0.75-1.98	0.653
AC	103 (50.5)	53 (51.5)	50 (48.5)				
CC	33 (16.2)	15 (45.5)	18 (54.5)				
IL-6							
rs1800796 (C-634G)							
CC	1 (0.5)	1 (100)	0 (0)	1.132	1.05	0.74-2.41	0.568
GC	30 (14.7)	15 (50)	15 (50)				
GG	172 (84.3)	82 (47.7)	90 (52.3)				
MTHFR							
rs1801133 (C667T)							
CC	76 (37.3)	36 (47.4)	40 (52.6)	0.004	1.02	0.575-1.80	0.952
CT	89 (43.6)	41 (46.1)	48 (53.9)				
TT	38 (18.6)	21 (55.3)	17 (44.7)				
rs1801131 (A-1298C)							
AA	107 (52.5)	54 (50.5)	53 (49.5)	0.435	0.83	0.478-1.442	0.509
AC	75 (36.8)	34 (45.3)	41 (54.7)				
CC	21 (10.3)	10 (47.6)	11 (52.4)				
SLCO1B1							
rs11045879 (C/T)							
CC	5 (2.5)	1 (20)	4 (80)	0.072	1.087	0.591-1.99	0.789
TC	53 (26.0)	26 (49.1)	27 (50.9)				
TT	146 (71.6)	71 (48.6)	75 (51.4)				
TNF-α							
rs1799724 (C-857T)							
CC	173 (84.8)	82 (47.4)	91 (52.6)	0.069	1.11	0.511-2.41	0.0792
CT	29 (14.2)	15 (51.7)	14 (48.3)				
TT	1 (0.5)	0 (0)	1 (100)				
rs1800629 (G-308A)							
AA	1 (0.5)	0 (0)	1 (100)	2.731	0.59	0.315-1.016	0.098
GA	53 (26.0)	31 (58.5)	22 (41.5)				
GG	149 (73.0)	66 (44.3)	83 (55.7)				
rs361525 (G-238A)							
GA	39 (19.1)	18 (46.2)	21 (53.8)	0.006	1.027	0.514-2.042	0.939
GG	164 (80.4)	79 (48.2)	85 (51.8)				
IL-10							
rs1800896 (T-1082C)							
CC	37 (18.1)	23 (62.2)	14 (37.8)	3.418	1.047	0.67-1.87	0.181

Supplementary table 2 (cont.). Genotype frequencies of single nucleotide polymorphisms in HCV infected PEG-IFN α -ribavirin therapy patients with and without SVR

	Study cohort n=204	SVR n= 98 (48 %)	Non-SVR n= 106 (52 %)				
TC	128 (62.7)	67 (52.3)	61 (47.7)				
TT	39 (19.1)	16 (41)	23 (59)				
IL-4							
rs2070874 (C-33T)							
CC	136 (66.7)	60 (44.1)	76 (55.9)	3.303	1.01	0.55-2.01	0.192
CT	59 (28.9)	33 (55.9)	26 (44.1)				
TT	9 (4.4)	5 (55.6)	4 (44.4)				
TNFRSF1B							
rs1061622 (T587G)							
GG	16 (7.8)	12 (75)	4 (25)	4.23	1.78	1.272-2.512	0.038
TG	76 (37.3)	40 (52.6)	36 (47.4)				
TT	112 (54.9)	46 (41.1)	66 (58.9)				
FCGR3A							
rs396991 (A559C)							
AA	88 (39.2)	41 (46.6)	47 (53.4)	2.024	1.93	0.77-4.88	0.155
AC	83 (36.8)	40 (48.2)	43 (51.8)				
CC	31 (11.3)	19 (61.3)	12 (38.7)				
FCGR2A							
rs1801274 (A519G)							
AA	54 (22.1)	29 (53.7)	25 (46.3)	0.228	0.847	0.428-1.676	0.633
AG	104 (46.6)	49 (47.1)	55 (52.9)				
GG	46 (18.1)	23 (40)	23 (50)				
STAT4							
rs7574865 (G/T)							
GG	110 (53.9)	57 (51.8)	53 (48.2)	1.697	0.693	0.398-1.204	0.193
GT	75 (36.8)	31 (41.3)	44 (58.7)				
TT	19 (9.3)	10 (52.6)	9 (47.4)				
PTPN22							
rs2476601 (C1858T)							
CT	17 (8.3)	7 (41.2)	10 (58.8)	0.35	1.354	0.494-3.709	0.554
CC	187 (91.7)	91 (48.7)	96 (51.3)				

Abbreviations: SNP, single nucleotide polymorphism; SVR, sustained virological response; OR, odds ratio; CI, confidence interval. Genotype frequencies were determined by χ^2 test using 3×2 or 2×2 tables as appropriate. Odds ratios and 95% CI per genotype were estimated by applying unconditional logistic regression. The P values less than 0.05 were considered significant.

SF1B/ IL-10/ IL-4 non-TTC was independently associated with SVR along with IL28B rs12979860 CC genotype, low baseline HCV RNA load and HCV genotypes 2 and 3 (Table 6). No independent association was detected for IL-(rs7248668, rs8105790, rs8099917) or TNFRSF1B (rs1061622), although the TNFRSF1B/IL-10/TNF α (-308) non-TTG was near to be significant (P=0.054).

Discussion

Different DAAs have been approved recently for HCV infection. These compounds used in different combinations, including PEG-INF-based triple therapies to INF-free oral bitherapies, have increased SVR rates even above 90%¹⁸. In this scenario, host genetics might beco-

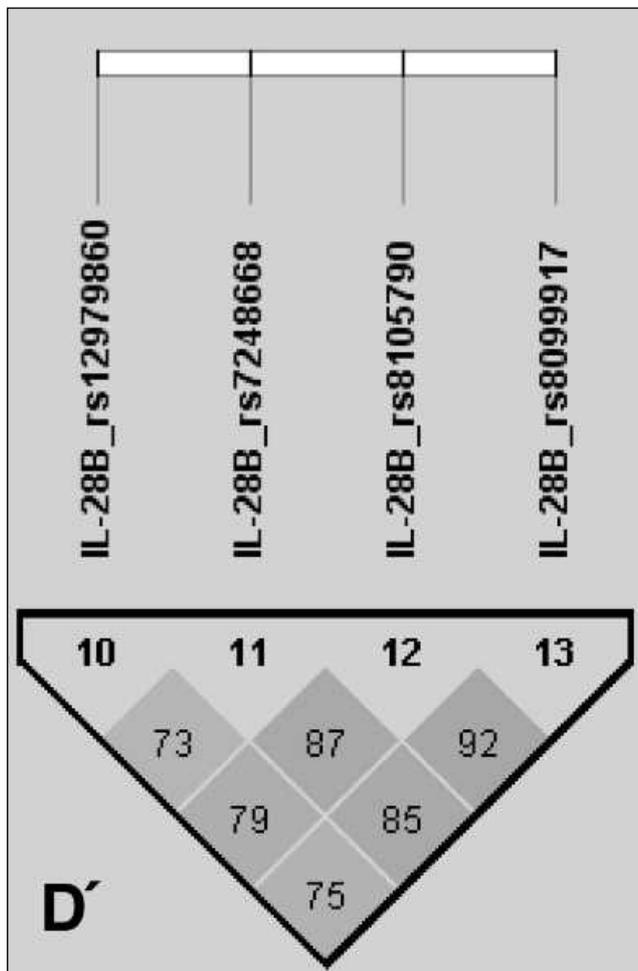


Figure 1. Pairwise linkage disequilibrium (D') pattern of IL28B region.

me appropriate to target patients who could more likely respond to conventional dual therapy with PEG-IFN and ribavirin which represents an efficient tool to individualize therapy.

The present work is a pharmacogenetic study conducted in a representative Spanish population of two cohorts of HCV infected patients who reached SVR and non-SVR to dual PEG-IFN and ribavirin therapy. In addition to well-known loci on the IL-28B gene, the previously unobserved SNPs located in the TNFRSF1B (rs1061622) and in different IL-10/ IL-4/ IL-6R/TNF α haplotypes were shown to have a significant association with the outcome of HCV treatment using PEG-IFN α and ribavirin. After multivariate analysis, the IL-28B rs12979860 CC genotype and TNFRSF1B/ IL-10/ IL-4 non-TTC haplotype were independently associated with SVR along with low baseline HCV RNA load and HCV genotypes 2 and 3 which may help to predict successful outcome in Spanish population.

Many studies over the last three decades have evaluated genetic predictors of HCV treatment response. Although the IL-28B genotype has emerged as the

strongest predictor of treatment response, a recent systematic review found that more than thirty genes have been associated with treatment response to INF-based therapies¹⁶. In this study we found an independent SVR predictive value for the IL-28B rs12979860 CC genotype but not for the rs7248668, rs8105790 or rs8099917 SNPs mainly because they are in linkage disequilibrium with rs12979860. It has been commented before the importance of the ethnicity to predict SVR. In fact, IL-28B frequency in each different SNP varies depending of the ethnicity which explains different responses to dual PEG-IFN/ribavirin treatment. Thus, for example, the Asian population shows a favorable 77% rs12979860 CC genotype distribution vs. 38% in Caucasian and 15% in African population which reflects the highest SVR of the Asian population^{13,19,20}. In addition, the most powerful IL-28B rs8099917 genotype predicts near to 90% of SVR in Asian population¹³ while in Caucasian population the best predictor was rs12979860 with a 60-70% of predictive value. Although Spanish population belongs to Caucasian ethnicity, to our knowledge there are no studies with a representative Spanish population from different regions which may add some variation to the current data. In this work we observed a 38.2% of rs12979860 CC genotype distribution in the general population studied, similar to the previously described in Caucasian population. Patients with SVR showed a 71.8% of rs12979860 CC genotype vs. 28.2% of non-responders, slightly lower than in general Caucasian population of the original studies¹³. Furthermore, the rs8099917 favorable TT genotype was the worst predictive IL-28B SNP. In terms of clinical use, the haplotype analysis revealed that combining all IL-28B SNPs results did not improve the strength of the association between the IL-28B genotype and SVR as compared with the results using rs12979860 genotype alone as recently was observed in other populations²¹. Cytokines and chemokines which are secreted by immune cells contribute to viral control, or liver damage. Examples are INF α/β , INF γ , TNF α , IL-6, IL-10, IL-4 and transforming growth factor beta (TGF β) between others²². They are able to render uninfected cells resistant to infection and cure the infected ones from the virus by stopping viral replication. In fact, PEG-INF-based therapy mimics the effect of host defense against HCV, increasing the expression of interferon stimulated genes (ISG) to kill HCV. In this regard, recent works have shown an association of different SNPs on TNF α (rs1800629), TGF β (rs1800469), IL-6 (rs1800797), IL-10 (rs1800896) and IL-4 (rs2070874) with SVR to PEG-INF ribavirin therapy^{16,23} which highlight the importance of an optimal host genetic anti-viral response for success of dual therapy.

TNF- α can bind to two different receptors: TNFRSF1A (tumour necrosis factor receptor superfamily 1A) and TNFRSF1B (tumour necrosis factor receptor superfamily 1B); which activates NF- κ B and triggers several inflammatory

Table 2. Genotype frequencies of single nucleotide polymorphisms in HCV infected PEG-IFN α -ribavirin therapy patients with and without SVR

Gene/ SNPs	Study cohort	SVR	Non-SVR	χ^2	OR	OR (95%CI)	P
	n=204	n= 98 (48 %)	n= 106 (52 %)				
	N (%)	N (%)	N (%)				
IL-28B							
rs12979860							
CC	78 (38.2)	56 (71.8)	22 (28.2)	26.47	2.27	1.57-3.27	8.75e-08
CT	97 (47.5)	38 (39.2)	59 (60.8)				
TT	29(13.7)	4 (14.3)	25 (85.7)				
<i>Allele of -response</i>							
T	(37.2)	(23.5)	(50)	30.55	4.75	2.57-8.76	3.25e-08
rs7248668							
CC	117 (52.0)	69 (59)	48 (41)	13.5	1.66	1.26-2.18	6.00e-04
CT	66 (30.9)	23 (34.8)	43 (65.2)				
TT	21 (9.8)	6 (28.6)	15 (71.4)				
<i>Allele of -response</i>							
T	(27.1)	(18.4)	35.2	14.61	3.04	1.66-5.54	1.00e-04
rs8105790							
TT	118 (52.2)	70 (59.3)	48 (40.7)	13.02	1.69	1.27-2.17	7.00e-04
TC	65 (30.7)	22 (33.8)	43 (66.2)				
CC	21 (9.8)	6 (28.5)	15 (71.4)				
<i>Allele of -response</i>							
C	(25.1)	(16.8)	(32.9)	13.83	3.04	1.68-5.49	2.00e-04
rs8099917							
TT	119 (58.3)	70 (58.8)	49 (41.2)	9.92	1.61	1.23-2.09	0.0016
TG	65 (31.4)	21 (32.3)	44 (67.7)				
GG	20 (9.3)	7 (35)	13(65)				
<i>Allele of -response</i>							
G	(24.9)	(17.9)	(31.4)	12.32	2.81	1.56-5.03	0.0016
TNFRSF1B							
rs1061622 (T587G)							
GG	16 (7.8)	12 (75)	4 (25)	4.23	1.78	1.27-2.51	0.038
TG	76 (37.3)	40 (52.6)	36 (47.4)				
TT	112 (54.9)	46 (41.1)	66 (58.9)				
<i>Allele of -response</i>							
T	(23.4)	(68.4)	(82.1)	4.22	1.85	1.31-2.42	0.041

Abbreviations: SNP, single nucleotide polymorphism; SVR, sustained virological response; OR, odds ratio; CI, confidence interval. Genotype frequencies were determined by χ^2 test using 3×2 or 2×2 tables as appropriate. Odds ratios and 95% CI per genotype were estimated by applying unconditional logistic regression. The P values less than 0.05 were considered significant.

Table 3. Genotype frequencies of single nucleotide polymorphisms in HCV infected PEG-IFN α -ribavirin treated patients who normalized serum hepatic enzyme values after 12 weeks of treatment.

Gene/ SNPs	AST < 35mg/dL N (%)	AST \geq 35mg/dL N (%)	χ^2	OR	OR (95%CI)	P
GNB3						
rs5443 (C825T)						
CC	51 (70.8)	21 (29.2)	6.34	2.14	1.24-3.54	0.042
CT	75 (78.1)	21 (21.9)				
TT	12 (52.2)	11 (47.8)				
IL-28B						
rs12979860						
CC	61 (81.3)	14 (18.7)	13.56	2.72	1.13-4.56	0.001
CT	65 (73)	24 (27)				
TT	12 (44.4)	15 (55.6)				
Gene/ SNPs	ALT < 45mg/dL N (%)	ALT \geq 45mg/dL N (%)	χ^2	OR	OR (95%CI)	P
GNB3						
rs5443 (C825T)						
CC	55 (69.6)	24 (30.4)	5.87	1.84	1.02-2.52	0.041
CT	63 (72.4)	24 (27.6)				
TT	14 (53.8)	12 (46.2)				
IL-28B						
rs12979860						
CC	63 (84)	12 (16)	13.25	2.91	1.41-6.04	0.001
CT	63 (70)	27 (30)				
TT	13 (48.1)	14 (51.9)				

Abbreviations: SNP, single nucleotide polymorphism; SVR, sustained virological response; OR, odds ratio; CI, confidence interval; AST: aspartate transaminase; ALT: alanine aminotransferase. Genotype frequencies were determined by χ^2 test using 3×2 or 2×2 tables as appropriate. Odds ratios and 95% CI per genotype were estimated by applying unconditional logistic regression. The P values less than 0.05 were considered significant.

Table 4. IL28B haplotype frequencies constructed with SNPs in HCV-infected groups (rs12979860, rs7248668, rs8105790, rs8099917)

Haplotype	Frequency	SVR (n=98)	Non-SVR (n=106)	χ^2	P value
CCTT	57	71 (139.2:56.8)	43.9 (92.2:117.8)	30.37	3.57e-08
TTCG	19.2	12.9 (25.2:170.8)	25.1 (52.7:157.3)	9.747	0.0018
TCTT	12.6	8.6 (16.8:179.2)	16.3 (34.3:175.7)	5.537	0.0186
CTCG	2.7	2.9 (5.7:190.3)	2.4 (5.1:204.9)	0.088	0.7669
TTTT	2.3	1 (1.9:194.1)	3.6 (7.5:202.5)	2.978	0.0844
TCCG	1.5	1 (2:194)	2 (4.2:205.8)	0.596	0.4402
CTTT	1.5	1.6 (3.1:192.9)	1.4 (3:207)	0.014	0.9062

Abbreviations: SVR, sustained virological response. The P-values were calculated from chi-squared (χ^2) test.

Table 5. Haplotype frequencies constructed with SNPs in HCV-infected groups (IL-10 rs1800896 (T/C); IL-4 rs2070874 (C/T); IL-6R rs2228145 (A/C); TNFRSF1B rs1061622 (T/G); TNF α rs1800629 (G-308A))

SNPs	Haplotype	Frequency	SVR (n=98)	Non-SVR (n=106)	χ^2	P
IL-10 (T/C)/ IL-4 (C/T)	TC	43.4	38 (121.5:74.5)	48 (108.2:101.8)	4.515	0.0335
IL-10 (T/C)/ IL-6R (A/C)	TA	29.3	24 (148.9:47.1)	34.3 (137.9:72.1)	5.192	0.0227
TNFRSF1B (T/G)/ IL-10 (T/C)	TT	36.6	30.5 (136.2:59.8)	42.3 (121.2:88.8)	6.064	0.0138
TNFRSF1B (T/G)/ TNF α (G-308A)	TG	67.6	62.4 (122.4:73.6)	72.4 (152.1:57.9)	4.61	0.0317
TNFRSF1B (T/G) / IL-10 (T/C) / TNF α (G-308A)	TTG	30.4	24 (148.9:47.1)	36.3 (133.7:76.3)	7.248	0.0071
TNFRSF1B (T/G)/ IL-10 (T/C)/ IL-4 (C/T)	TTC	32.5	25.9 (145.1:50.9)	38.6 (129:81)	7.361	0.0067

Abbreviations: SVR, sustained virological response. The P-values were calculated from chi-squared (χ^2) test.

Table 6. Independent predictors of SVR in patients who completed the treatment regimen with PEG-IFN α and ribavirin

Parameter	OR (95% CI)	P value
Age.years (≤ 48 vs. > 48)	1.1 (0.87-1.3)	0.09
HCV genotype (2. 3 vs. 1a. 1b. 4)	3.61 (1.98-7.25)	0.0002
Fibrosis (F0-F2 vs. F3-F4)	2.61 (0.97-8.54)	0.145
Baseline HCV-RNA (log UI/mL; ≤ 5.79 vs. > 5.79)	3.54 (2.1-10.41)	0.014
LDL cholesterol (mg/dL; ≥ 110 vs. < 110)	2.04 (1.41-7.98)	0.071
IL-28B rs12979860 (CC vs. CT/ TT)	2.11 (1.1-5.48)	0.004
TNFRSF1B rs1061622 (GG vs. TG/ TT)	1.55 (0.84-9.47)	0.157
TNFRSF1B/ IL-10 / TNF α (-308); Non-TTG vs. TTG	1.98 (0.9-6.51)	0.054
TNFRSF1B/ IL-10/ IL-4; Non-TTC vs. TTC	2.01 (1.1-5.87)	0.047

Abbreviations: SVR, sustained virological response; OR, odds ratio; CI, confidence interval; pegIFN α : pegylated interferon alpha. OR of having a SVR to PEG-IFN α -ribavirin therapy were calculated. P values less than 0.05 were considered statistically significant.

pathways with anti-viral properties^{24,25}. Rs1061622 is located in the exon 6 of TNFRSF1B, and the change of T to G causes a functional amino acid substitution at codon 196 from methionine (Met) to arginine (Arg) in the translated protein TNF receptor 2 (TNFR2). This variant G is supposed to produce a change of biological function of the TNFR2 protein affecting TNF- α binding and NF- κ B activation which consequently modifies INF I and III production²⁵. Interestingly, in some drug sensitivity studies, rs1061622 was found to be associated with a favorable prognosis of non-small cell lung cancer patients treated with chemoradiotherapy²⁶ and with a beneficial response to infliximab in Crohn's disease²⁷. In this work, it was observed for the first time an association of the TNFRSF1B rs1061622 G allele with a SVR. In the same way, different haplotypes combining IL-10, IL-6R, TNF α and IL-4 cytokines increased the predictive value of TNFRSF1B rs1061622, indicating that anti-viral inflammatory response is a complex molecular process which encompasses a broad number of inflammatory cells and molecular pathways. In fact, TNFRSF1B/ IL-10/ IL-4 non-TTC

haplotype reached significant association with SVR in the multivariate analysis.

As part of the different inflammatory pathways involved in HCV liver inflammation, GNB3 rs5443 CC genotype, which encodes a splice variant of the β 3 subunit of heterotrimeric G proteins, has been associated with an inactivation of GNB3, thus reducing the activation of broad number of inflammatory receptors coupled to GNB3. This fact, may explain rapid normalization of AST and ALT hepatic enzymes in GNB3 CC carriers observed in this work, although no association with SVR was observed.

However, as we commented before, host genetics cannot explain the 100% of SVR. There are other clinical and viral factors that influence the response to PEG-IFN/ ribavirin treatment. In this work we observed an association of lower age, lower basal HCV-RNA load, HCV2 and HCV3 genotypes, lower fibrosis stage F0-F2, and elevated LDL cholesterol with a SVR, which confirm previous reports^{28,29}. However, the multivariate analysis independently associated lower basal HCV-RNA load, HCV 2 and

3 genotypes as predictors of SVR, probably because the study sample did not reach enough size.

In summary, we found genetic, viral and clinical parameters associated with SVR, particularly IL-28B rs12979860 CC, TNFRSF1B/IL-10/IL-4 non-TTC haplotype, low baseline HCV RNA load and HCV genotypes 2 and 3 which may help to predict successful outcome to PEG-IFN/ribavirin therapy in Spanish population. Further analysis would be required to confirm these associations with triple therapy including new DAAs.

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